Comparison of the capacity of biological desulfurization of *Thiobacillus ferrooxidans* from different sulfur-containing substrates with or without additional ferrous iron

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Abstract: *Thiobacillus ferrooxidans*, abbreviated as *T. ferrooxidans* is one of the important microorganisms in the field of biological desulfurization. Effects of ferrous iron and sulfur-containing substrates on biological desulfurization of *T. ferrooxidans* were studied. Results show that in the absence of Fe²⁺, *T. ferrooxidans* can utilize three kinds of sulfur-containing substrates of Na₂S₂O₃, elemental S and Na₂SO₃ for growth and metabolism. For utilization complexity, Na₂S₂O₃ was easiest to use, next was elemental S, and Na₂SO₃ was the worst for use. During the utilization of ferrous iron and sulfur-containing substrates by *T. ferrooxidans*, the iron oxidation system was first started. With the decrease of the Fe²⁺ concentration, the sulfur oxidation system was started, and then the two systems synergistically acted. The presence of three sulfur-containing substrates had different effects on Fe²⁺, especially the inhibition of Na₂SO₃ was significant, and complete oxidation of ferrous iron needed more time. The isolated *T. ferrooxidans* is applied to the removal of H₂S gas, aiming to provide a new technological approach for biological removal of H₂S.

Keywords: *Thiobacillus ferrooxidans*, ferrous iron, sulfur-containing substrate, biological desulfurization **DOI:** 10.25165/j.ijabe.20241703.8613

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1 Introduction

With the rapid development of modern agriculture, the production of organic solid waste such as straw and livestock manure has been increasing year by year. Improper treatment and disposal can cause serious environmental pollution problems, endanger human health, and also result in serious resource waste. Anaerobic fermentation is a process in which insoluble organic matter is gradually decomposed into water-soluble small molecule organic matter through the metabolic activity of facultative and anaerobic microorganisms in an anaerobic environment, ultimately producing biogas mainly composed of methane. It is an effective way to achieve the resource utilization and energy recovery of agricultural waste such as straw and livestock manure. The biogas produced by anaerobic fermentation contains H₂S, which has extremely strong toxicity^[1-3]. When using biogas, H₂S removal treatment must be carried out first^[4-6]. Recently, as a new desulfurization technology, H₂S removal by T. ferrooxidans has gained much attention^[7-9]. T. ferrooxidans is a kind of obligate aerobic and eosinophilic chemoautotrophic bacterium^[10-12]. It exists by oxidizing Fe²⁺ into Fe³⁺ and by oxidizing low valent sulfur into

sulfate, and is gram negative^[13,14]. The removal of H₂S by *T*. *ferrooxidans* can utilize its indirect oxidation^[15-17]. The desulfurization mechanism is^[18]:

$$H_2S + Fe_2(SO_4)_3 \rightarrow 2FeSO_4 + H_2SO_4 + S \downarrow$$
(1)

$$2FeSO_4 + 0.5O_2 + H_2SO_4 \xrightarrow{T.ferrooxidans} H_2O + Fe_2(SO_4)_3$$
(2)

The rate of Fe²⁺ oxidation by *T. ferrooxidans* is at least 200 000 times faster than the chemical oxidation rate $alone^{[19]}$, but with the increase of Fe³⁺, a certain amount of ferro-vanadium precipitation will be produced in the medium^[20,21]. It can also be used to oxidize H₂S to elemental S and H₂SO₄ by its direct oxidation^[22,23]. The reaction formula is:

$$2H_2S + O_2 \rightarrow 2S + 2H_2O \tag{3}$$

$$2S + 3O_2 + 2H_2O \rightarrow 2H_2SO_4 \tag{4}$$

With comparison, *T. ferrooxidans* tends to use ferrous iron more, and most researches on *T. ferrooxidans* use ferrous iron as the energy source^[24-26]. In order to avoid the precipitation during the indirect oxidation process of *T. ferrooxidans*, improve its direct oxidation of low valent sulfur^[27-29], and realize better application for H₂S removal, this paper studied the utilization of sulfur-containing substrates by *T. ferrooxidans* in the presence and absence of Fe²⁺, providing the theoretical basis for its industrialization application in biological desulfurization.

2 Materials and methods

2.1 Materials

- 2.1.1 Microbial strain and medium
 - T. ferrooxidans was isolated from acid mine water by dilution

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and coating method, and was identified by colony morphology, microscopic observation, staining treatment, culture characteristics and 16S rRNA gene sequence analysis.

1) Isolation of strains

(1) Method 1: Serial dilution method

100 μ L culture medium was inoculated in 1 mL 9K liquid medium (15 mL test tube), and 10 tubes of gradient dilution (10-1) were cultured at 30°C, 170 r/min, and the color change of test tubes was observed day by day for 10 days. In addition, single colony isolation was performed on 9K solid medium.

9K liquid medium: $(NH_4)_2SO_4$ 3.00 g, K_2HPO_4 0.50 g, KCL 0.10 g, MgSO₄·7H₂O 0.50 g, Ca(NO₃)₂ 0.01 g, FeSO₄·7H₂O 44.30 g, distilled water 1000 mL, pH 2.0.

9K solid medium: adding 2% agarose to 9K liquid medium.

(2) Method 2: Coating method

Since many bacteria that can oxidize ferrous ions also have sulfur oxide function, it is speculated that the bacteria in the culture medium may also oxidize low-price sulfur. 200 μ L culture medium was directly coated on SO (sulfur oxidization medium) solid medium and cultured at 30°C for 2 d. Individual colonies were selected and striated on the same medium for purification.

SO liquid medium: CH₃COCOONa 1.98 g, Na₂S 0.21 g, yeast extract 0.3 g, distilled water 300 mL, pH 7.0.

SO solid medium: adding 2% agar powder to SO liquid medium.

2) Identification of strains

According to the colony morphology, microscopic observation, staining treatment, culture characteristics and 16S rRNA sequence analysis, the strains were determined. The primers used for 16S gene (5'rRNA sequence analysis were 8f (5'-AGAGTTTGATCMTGGC-2') and 1542r AAAGGAGGTGATCCA-2'). PCR reaction conditions were as follows: denaturation temperature 94°C, 2 min; annealing temperature 55°C, 1 min; extended temperature 72°C, 1 min, total 25 cycles.

2.1.2 Reagent

(NH₄)₂SO₄, AR, Guangdong Guanghua Chemical Co., Ltd; K₂HPO₄, AR, Nanjing Chemical Reagent Factory; KCl, AR, Nanjing Chemical Reagent Factory; MgSO₄·7H₂O, AR, Shanghai Tongya Fine Chemical Factory; Ca(NO₃)₂, AR, Wenzhou Chemical Factory; FeSO₄·7H₂O, AR, Sinopharm Chemical Reagent Co., Ltd.; CH₃COCOONa, AR, Sinopharm Chemical Reagent Co., Ltd.; Na₂S, AR, Sinopharm Chemical Reagent Co., Ltd.; Yeast extract, AR, Shanghai Shenggong Biotechnology Co., Ltd. S, AR, Nanjing Ningshi Chemical Reagent Co., Ltd.; Na₂S₂O₃, AR, R&D center of Guangdong Chemical Reagent Engineering Technology; Na₂SO₃, AR, Nanjing Ningshi Chemical Reagent Co., Ltd.

2.1.3 Apparatus

Air oscillating shaker (THZ-C constant-temperature oscillator, Taicang Jiangsu); pH meter (6171C type microcomputer pH/ mV/temperature, Shanghai Jenco Electronic Co., Ltd.); scales (Sartorius electronic scales, Beijing Sartorius scale Co., Ltd.).

2.2 Methods

9K liquid medium containing no Fe^{2*} was prepared, and elemental S, $Na_2S_2O_3$ and Na_2SO_3 were added separately. Measured as per S, concentration of S added was 1 g, which was 0.03 mol/L, cultured at 30°C and 120 r/min. Due to that sulfuric acid generation by oxidation of sulfur compounds is an acid-producing process, and the degree of pH reduction of the solution can be used to indicate the amount of sulfur compounds oxidized by the bacteria. Therefore, the pH and SO_4^{2-} concentration in the solution were measured at regular intervals, and utilization of sulfur compounds by *T. ferrooxidans* was analyzed.

In order to investigate the utilization of two energy sources by *T. ferrooxidans* in the simultaneous presence of ferrous and sulfurcontaining substrates, 9K liquid medium containing Fe²⁺ was prepared, and elemental S, Na₂S₂O₃ and Na₂SO₃ were added separately. Measured as per S, concentration of S added was 1 g/L, which was 0.03 mol/L, cultured at 30°C and 120 r/min. PH, Fe²⁺ and SO₄²⁻ concentration in the solution were measured at regular intervals, and effects of ferrous iron and sulfur-containing substrates on biological oxidation process of *T. ferrooxidans* was analyzed.

The pH value was determined by pH meter; Fe^{2+} determined by O-phenanthroline spectrophotometry^[30]; SO_4^{2-} concentration determined by barium chromate spectrophotometry^[31].

3 Results and discussion

The change of pH in solution is shown in Figure 1 when T. ferrooxidans uses three sulfur-containing substrates as energy materials with or without additional ferrous iron, and the change of SO_4^{2-} concentration in six culture systems is shown in Figure 2. It was found that among the three sulfur-containing substrates, Na₂S₂O₃ was easiest to use, and next was elemental S, and Na₂SO₃ was worst to use. When there were only sulfur-containing substrates in the medium without Fe2+, T. ferrooxidans had a certain lag period at the beginning, with different duration time. During the lag period, decrease of pH of the solution was not significant, and the concentration of SO₄²⁻ in the solution was very low. With the growth of T. ferrooxidans, the pH value in the solution decreased, and the concentration of SO_4^{2-} in the solution also gradually increased and then stabilized. Since Na2SO3 is a weak acid and strong alkali salt, the pH of the solution during the whole process of T. ferrooxidans growth with Na₂SO₃ as substrate is higher than that of elemental S and $Na_2S_2O_3$. Oxidation of sulfur compounds by T. ferrooxidans is an acid-producing process, and the pH value in the solution still shows a downward trend.



Figure 1 The pH change of the solution with different substrates

Though Na₂S₂O₃ would become disproportionate as the SO₃²⁻ and S under acidic conditions, the growth rate of SO₄²⁻ was still faster than that when Na₂SO₃ was used as a substrate. So it can be inferred that the oxidation of S₂O₃²⁻ and SO₃²⁻ by *T. ferrooxidans* is not the effect by the same enzyme, and the utilization rate of S₂O₃²⁻ is faster than that of SO₃^{2-[12,3]}. Since SO₃²⁻ in Na₂SO₃ is of in the ionic state and the air shall oxidize part of the SO₃²⁻, the concentration of SO₄²⁻ in the solution containing Na₂SO₃ as substrate in the first 2 days thus was higher than that in the solution containing elemental S as substrate. But with the utilization of elemental S by *T. ferrooxidans*, the concentration of SO₄²⁻ in solution containing elemental S shall increase rapidly. At the end of the experiment, the concentration of SO_4^{2-} in the solution with elemental S as the substrate increased by 105%, the concentration of SO_4^{2-} in the solution with $Na_2S_2O_3$ as the substrate increased by 112%, and the concentration of SO_4^{2-} in the solution with Na₂SO₃ as the substrate increased by 72% with the smallest increase. For T. ferrooxidans growth with elemental S as the substrate, thallus was first adsorbed on the surface of elemental S particles, and elemental S passed through the cell wall and entered the interior of the cell to form a polysulfide compound with glutathione, then the first-grade product sulfite was produced. Then SO₃²⁻ was further oxidized into SO_4^{2-} by action of adenosine monophosphate. The whole process involved a series of complex mass transfer processes such as adsorption of T. ferrooxidans on the surface of solid particles and diffusion of products through the cell wall. Due to the slow adsorption rate of T. ferrooxidans on the surface of elemental S particles, the step of solid-phase interface mass transfer became a rate-limiting step for the utilization process of elemental S^[34,35]. Once the sulfur was oxidized to SO_3^{2-} , energy utilization by cells became faster.



Figure 2 The SO_4^{2-} concentration change of the solution with different substrates

The change of Fe^{2+} concentration in the solution with Fe^{2+} is showed in Figure 3. Since Fe^{2+} oxidation by *T. ferrooxidans* is easier, and elemental S is a poorly soluble solid particle, there was no adaptation period of bacteria during the cultivation process, that is, the utilization of Fe2+ was not limited by presence of S, and the Fe²⁺ enzyme system was first activated^[36,37]. Na₂S₂O₃ and Na₂SO₃ had some inhibition on the oxidation of Fe2+, especially the inhibition of Na₂SO₃ was more obvious. And the time needed for complete oxidation of ferrous iron was longer, for 6 days. While in the solution added with Na₂S₂O₃, ferrous iron had been completely oxidized in 2 d. With the growth of bacteria, the concentration of Fe²⁺ in the solution gradually decreased until it was completely oxidized, the energy utilization metabolic pathway of sulfur-oxidase started, and the pH value in the solution began to decrease significantly. The change of pH value in the solution of T. ferrooxidans with FeSO₄ and Na₂S₂O₃ as substrates was the same as that with FeSO₄ and S, and with a trend of increase first and then decrease, while the former's rangeability was bigger than that of the latter. For T. ferrooxidans with FeSO₄ and Na₂S₂O₃ as substrates, the pH value in the solution showed a downward trend. Though the addition of Na₂SO₃ led to an increase in the initial pH value of the solution, the pH in the solution dropped rapidly after 1 day. The reason may be that Fe^{2+} in the solution is first oxidized by T. *ferrooxidans* and results in Fe^{3+} which can oxidize SO_3^{2-} , thus decreasing the pH of the solution. Moreover, with the growth of T. *ferrooxidans*, the enzyme system that oxidizes SO_3^{2-} begins to take effect, thus accelerating the decrease of the pH.



Figure 3 The Fe^{2+} concentration change of the solution with different substrates

Due to the presence of energy substance Fe²⁺, the concentration of SO_4^{2-} in the solution with Fe^{2+} all increased to some extent. Moreover Fe³⁺ oxidized by *T. ferrooxidans* from Fe²⁺ had an oxidation effect, thus, the utilization of intermediate product of sulfur compound metabolite by T. ferrooxidans was better than that in the absence of Fe^{2+} . Due to oxidation of SO_3^{2-} by air, the SO_4^{2-} concentration in the solution with FeSO₄ and Na₂SO₃ as substrates in the first 4 d was higher than that in the solution with FeSO₄ and S as substrates. However, in the later stage of culture, SO_4^{2-} in the latter solution increased rapidly with the increase of the amount of T. ferrooxidans adsorbed on elemental S. In solutions with FeSO₄ and S, FeSO₄ and Na₂S₂O₃, and FeSO₄ and Na₂SO₃ as substrates separately, maximum concentration of SO₄²⁻ were 18.48 g/L, 18.80 g/L, 18.32 g/L separately, which, in relative to the increase amplitude of the initial value, were higher than that with absence of Fe^{2+} . At the later stage of the experiment, SO_4^{2-} concentration in the three solutions showed a downward trend due to hydrolysis and precipitation of some Fe3+ in the solution.

4 Conclusions

In the absence of Fe^{2+} , $Na_2S_2O_3$ was easiest to use for T. ferrooxidans, next was elemental S, and Na₂SO₃ was difficult to use. When ferrous iron and sulfur-containing substrates were present together, the iron oxidation system of T. ferrooxidans was first started. Then these two oxidation systems synergistically acted. Elemental S did not inhibit the oxidation of Fe²⁺, while Na₂S₂O₃ and Na₂SO₃ had certain inhibition on the oxidation of Fe²⁺, especially the inhibition of Na₂SO₃ was more obvious. Due to its special physiological and metabolic characteristics, T. ferrooxidans has become one of the most important strains in the fields of biological desulfurization of fuel and acid waste gas, biological hydrometallurgy, acid mine drainage treatment and so on^[38,39]. Its research not only provides a new way for the biological removal of H₂S, but also expands the research and application of biotechnology in the field of waste gas treatment, which is of great significance for the protection of ecological environment.

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